

Porwenins A and B, New Clerodane Diterpenoids from *Portulaca okinawensis*

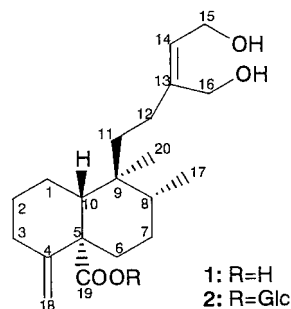
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Two new clerodane-type diterpenes, porwenins A (**1**) and B (**2**), were isolated from *Portulaca okinawensis*, and the structures were elucidated by spectroscopic data.

From biosynthetic and chemosystematic points of view, we have studied the structures of diterpenoid constituents of *Portulaca* species.¹ Among them, *Portulaca okinawensis* (Portulacaceae) is a rare, peculiar plant, distributed on rock cliffs on the seashore of Okinawa Island. Recent investigation of extracts of *P. okinawensis* resulted in the isolation of two new clerodane diterpenes, porwenins A (**1**) and B (**2**). Here we describe the isolation and structure elucidation of **1** and **2**.



The molecular formula, C₂₀H₃₂O₄, of porwenin A (**1**) was established by HRFABMS. The IR spectrum suggested the presence of hydroxy (3348 cm⁻¹) and carboxylic (1710 cm⁻¹) groups. The gross structure of **1** was deduced from detailed analysis of the ¹H and ¹³C NMR data (Table 1) aided by 2D NMR experiments (¹H–¹H COSY, ¹³C–¹H COSY, and HMBC). The ¹H and ¹³C NMR data indicated that compound **1** possessed one carboxyl group, one trisubstituted olefin, one exomethylene, two hydroxy methylenes, two methyl groups, two sp³ quaternary carbons, two methines, and seven methylenes. Since three of five unsaturations were accounted for, it was indicated that **1** contained two rings. The ¹H–¹H COSY spectrum revealed connectivities of C-1 to C-3 and C-10, C-6 to C-8 and C-17, and C-11 to C-12 (Figure 1). HMBC correlations (Figure 1) of H-3 to C-4, H-18 to C-3, C-4, C-5, and C-19 (δ_c 178.1), and H-10 to C-1 and C-19 suggested the presence of a cyclohexane ring (ring A), in which an exomethylene unit and a carboxyl group were attached to C-4 and C-5, respectively. The presence of another cyclohexane ring (ring B) with Me-17 and Me-20 at C-8 and C-9, respectively, was elucidated by HMBC correlations of H-6 to C-10 and C-8, H₃-17 to C-7, C-8, and C-9, and H₃-20 to C-9 and C-10. The side chain,

Table 1. ¹H and ¹³C NMR Data for Porwenins A (**1**) and B (**2**) in CD₃OD^a

position	¹ H		¹³ C	
	1	2	1	2
1	1.63m, 2.30m	1.63m, 2.32m	22.6	22.5
2	1.93m	1.48m, 1.78m	29.0	29.6
3	2.17brd(13.4), 2.56m	2.15brd(13.4), 2.49dt(13.4,4.9)	34.1	34.0
4			153.6	152.6
5			52.3	52.9
6	1.30m, 2.51m	1.38m, 2.58dt(12.8,3.0)	36.1	35.6
7	1.50m	1.33m, 1.93m	29.9	29.1
8	1.58m	1.58m	38.2	38.2
9			40.9	41.0
10	1.33dd (12.8, 3.1)	1.36dd(12.8, 3.0)	52.6	53.2
11	1.46m, 1.56m	1.42m, 1.54m	37.6	37.6
12	1.88m, 2.24m	1.85m, 2.04m	28.3	28.4
13			143.6	143.6
14	5.45t (6.7)	5.45t (6.7)	127.3	127.3
15	4.13d (6.7)	4.13d(6.7)	58.8	58.8
16	4.09s	4.08s	60.3	60.2
17	0.82d (6.1)	0.82d(6.7)	16.3	16.3
18	4.76brs, 4.78brs ^b	4.85brs, 4.86brs	109.7	110.3
19			178.1	174.1
20	0.75s	0.74s	18.0	18.6
1'		5.45d(7.4)		95.6
2'		3.33m		74.2
3'		3.32m		78.8
4'		3.35m		71.0
5'		3.39m		78.6
6'		3.67dd(12.2,1.2), 3.80dd(12.2,4.3)		62.3

^a 500 MHz for ¹H and 125 MHz for ¹³C; *J* values (Hz) in parentheses. ^b The spectrum was measured in CD₃COCD₃.

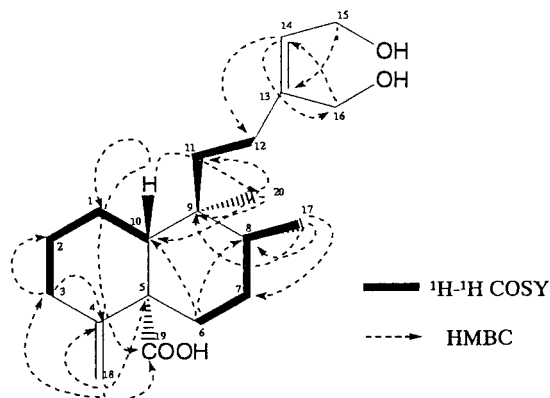


Figure 1. Selected 2D NMR correlations of porwenin A (**1**).

a 1,4-dihydroxy-2-buten-2-ylethyl group, was attached to C-9 due to HMBC cross-peaks of H₃-20 to C-9 and C-11.

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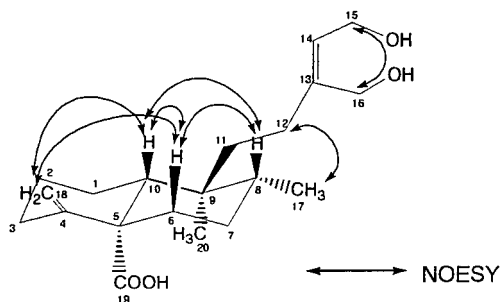


Figure 2. Selected NOESY correlations of porwenin A (**1**).

NOESY correlations (Figure 2) of H₂-18 to H-10, H-6b, H-10 to H-6b, H-8, and H-6b to H-8 indicated chair conformations of rings A and B, a *trans* relationship between rings A and B, α -orientations of Me-17, Me-20, and a carboxyl group at C-5, and β -orientation of H-10. Therefore, the structure and relative stereochemistry of **1** were assigned as shown.

The molecular formula, C₂₆H₄₂O₉, of porwenin B (**2**) was established by HRFABMS. The IR spectrum indicated the presence of hydroxy (3356 cm⁻¹) and ester (1734 cm⁻¹) groups. The ¹H and ¹³C NMR spectra of **2** were similar to those of porwenin A (**1**). Analysis of the ¹H and ¹³C NMR data (Table 1) and the ¹³C-¹H COSY spectrum of **2** provided five methines (all of which were bearing an oxygen atom) and one hydroxymethylene in addition to all the carbons of **1**. ¹H-¹H connectivities of C-1' to C-6' and the ¹H and ¹³C chemical shifts (Table 1) of **2** suggested the presence of a glucose moiety. Furthermore, the anomeric proton signal at δ_{H} 5.45 (d, $J = 7.39$, H-1') and the carbon signal at δ_{C} 95.6 (C-1') having the ¹J_{CH} value of 163 Hz, which was deduced from INEPT-nondecoupling measurement,² revealed the β -configuration of the glycoside bond at C-1'. Comparison of the ¹³C NMR data of **1** and **2** and the HMBC cross-peak of H-1' to C-19 in **2** indicated that the 19-carboxyl group of **1** was glycosylated in **2**. Thus, the structure of porwenin B was assigned as **2**. The relative stereochemistry of **2** was elucidated to be the same as that of **1** by NOESY correlations (Figure 2).

Porwenins A (**1**) and B (**2**) possess in common a 1,4-dihydroxy-2-buten-2-ylethyl group at C-9, which is characteristic of the diterpenoids found in *Portulaca* plants, while the ring skeleton of both **1** and **2** is that of normal-type clerodane diterpenes without skeletal rearrangement such as the major constituents of *Portulaca* cv. Jewel.³

Experimental Section

General Experimental Procedures. IR spectra were obtained with a JASCO infrared microscope and Jansenn MFT-2000 spectrometer. Optical rotations were obtained with a JASCO DIP-370 polarimeter. HRFABMS measurements were carried out using a JEOL HX-110 mass spectrometer. ¹H and ¹³C spectra, including 2D experiments, were measured using a JEOL α -500 NMR spectrometer. TMS and the resonance of residual CD₃OD (49.0 ppm) were used as internal reference for ¹H and ¹³C NMR, respectively.

Plant Material. *P. okinawensis* was collected on the rock cliff of the seashore at Yomitan Village, Okinawa Island, in September 1999 and cultivated in Yokohama. This plant was identified by Dr. T. Shinzato (Faculty of Agriculture, University of the Ryukyus). A voucher specimen has been deposited at the Institute of Biomaterials and Bioengineering, Tokyo Medical and Dental University.

Extraction and Separation. The fresh whole plant (133.2 g) of *P. okinawensis* was extracted with MeOH, and the extract (4.2 g) was partitioned between hexane and 90% aqueous MeOH. The MeOH-soluble portion was partitioned with EtOAc and H₂O. The EtOAc-soluble materials were subjected to an ODS column (YMC-GEL, ODS-AM 120-S50, YMC co. LTD, Japan) with CH₃CN-H₂O (0:100 \rightarrow 100:0) to give six fractions. The fraction eluted with CH₃CN-H₂O (40:60) was separated with CH₃CN-H₂O (20:80 \rightarrow 80:20) by C₁₈ HPLC (Capcell pak, UG-120, 3.0 \times 250 mm, Shiseido; flow rate, 8 mL/min; detection, 210 nm) to afford porwenins A (**1**, 4.7 mg) and B (**2**, 7.5 mg).

Porwenin A (1): colorless amorphous solid; $[\alpha]_{\text{D}}^{23} +45.5^{\circ}$ (c 0.24, MeOH); IR (neat) ν_{max} 3348, 1710 cm⁻¹; ¹H and ¹³C NMR (Table 1); HRFABMS (negative) m/z 335.2247 (M - H)⁻ (calcd for C₂₀H₃₁O₄, 335.2222).

Porwenin B (2): colorless amorphous solid; $[\alpha]_{\text{D}}^{23} +36.6^{\circ}$ (c 0.37, MeOH); IR (neat) ν_{max} 3356, 1734 cm⁻¹; ¹H and ¹³C NMR (Table 1); HRFABMS (positive) m/z 521.2712 (M + Na)⁺ (calcd for C₂₆H₄₂O₉Na, 521.2727).

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References and Notes

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